

# ANTHELMINTIC RESISTANCE IN SMALL RUMINANT FARMS: AN ONGOING CHALLENGE FOR PERAK FARMERS TO CONTROL HELMINTHS

PREMAALATHA B.<sup>1\*</sup>, CHANDRAWATHANI P.<sup>1</sup>, ERWANAS A.I.<sup>1</sup>, LILY ROZITA M.H.<sup>1</sup>, JAMNAH O.<sup>1</sup>, AIZAN Y.<sup>2</sup> AND RAMLAN M<sup>1</sup>

<sup>1</sup> Veterinary Research Institute, 59 Jalan Sultan Azlan Shah, 31400 Ipoh, Perak .

<sup>2</sup> Perak State Department of Veterinary Services, Jalan Sultan Azlan Shah (U), 31400 Ipoh, Perak.

\* Corresponding author: princess\_latha2280@yahoo.com

**ABSTRACT.** This study was done to evaluate the status of resistance to nematode populations on four major groups of anthelmintics. Nematode faecal egg count reduction tests (FECRT) were conducted on six free grazing small holder goat farms (namely Farms A-F) in Perak. The four drug groups tested in this study were Benzimidazoles, Imidazothiazoles, Macrocytic Lactones and Salicylanilides. Faecal samples were subjected to the standard procedures of McMaster for worm egg estimation and also larval cultures for third stage larvae identification. Results of Farm A, Farm C and Farm D showed resistance to all 4 drugs groups of anthelmintic that were tested and the major worm population of this farm were *Haemonchus contortus* (49.75%), *Trichostrongylus colubriformis* (47.71%) and *Cooperia* sp. (2.53%). The results of Farm B and Farm F showed resistance to all the 3 groups of anthelmintics except Levamisole and the worm population of this farm was *Haemonchus contortus* (47.30%), *Trichostrongylus* sp. (38.44%), *Oesophagostomum columbianum* (13.50%) and *Cooperia* sp. (0.76%) Farm E was resistant to Benzimidazoles and

suspected resistance to 3 drugs groups; Levamisole, Macrocytic Lactones and Closantel. The worm population for Farm E was *Haemonchus contortus* (71.35%), *Trichostrongylus* sp. (27.6%) and *Oesophagostomum* sp. (2%). These farms have a history of using the same anthelmintics over the past few years, thus leading to the severe case of anthelmintic resistance. Recommendations have been made to control helminths using alternative approaches such as cut and carry feeding, herbal medication using Neem leaves (*Azadirachta indica*) and rotational grazing as well as improvement in management of animals to increase their immunity.

*Keywords:* FECRT, anthelmintic resistance, free grazing small holder

## INTRODUCTION

Helminth infection has been recognised as the most important cause of mortality and morbidity in small ruminants in Malaysia (Fatimah *et al.*, 1985; Sani and Rajamanickam, 1990; Sani *et al.*, 2004). Anthelmintic resistance in sheep and goats in Malaysia was reported in the early 1990s, with demonstrations of resistance to

the benzimidazoles (Rahman, 1993;1994; Pandey and Sivaraj, 1994), levamisole (Dorny *et al.*, 1994) and the first case of multiple anthelmintic resistance reported by Sivaraj and co-worker in 1994 (Sivaraj *et al.*,1994).

In 2006 FECRT was conducted in 18 small ruminants private farms located in Peninsular Malaysia and it was found that nematode populations on all farms showed resistance to salicylanilides closantel while 13 farms showed resistance to oxfendazole, 8 farms showed resistance to imidazothiazoles and 4 farms showed resistance to moxidectin (Khadijah *et al.*, 2006).

In 2012, two goat farms experiencing severe mortality up to 30% in the flock were tested for anthelmintic resistance using the Faecal Egg Count Reduction Test (FECRT) with four anthelmintics from the group; Benzimidazoles, Imidazothiazoles, Macrocytic Lactones and Salicylanilides. Results showed that *Haemonchus contortus* and *Trichostrongylus colubriformis* in these two farms were resistant to all the four anthelmintic groups tested (Chandrawathani *et al.*, 2013).

In view of the escalating anthelmintic resistance problem in small ruminants in Malaysia, the Veterinary Research Institute has embarked on a programme to identify farms with severe helminthiasis whereby common anthelmintics seem ineffective and thereon establish a database of regions or areas which coincide with different anthelmintic resistance status. This is an ongoing study conducted on six free grazing

small holder goat farms in several districts of Perak. The farms were investigated over a period of a year (January to December 2012). The objective of this study was to evaluate the current status of nematode anthelmintic resistance in these farms, as these farms experienced severe mortality and morbidity due to helminthiasis. From this study the farmers would be better able to control helminthiasis by acknowledging suitable and effective drugs and in the long term make their livestock venture profitable.

## MATERIALS AND METHODS

The Faecal Egg Count Reduction Test (FECRT) was conducted on 6 goat farms near Ipoh, Perak. Four types of anthelmintics, Benzimidazoles (Albendazole Vetpharm Laboratories (S) Pte. Ltd.), Imidazothiazoles (Levamisole, Bovet Pharmaceutical Ltd.), Salicylanilides (Closantel, Janssen Pharmaceutical N.V.) and Macrocytic Lactones (Ivermectin, Man./Fab: KELA N.V.) were used. Animals were divided to 5 equal groups of which, 4 are treated groups and 1 control group. Each group of 6-10 animals were treated according to the manufacturers recommended dose rates based on individual body weight and the control group were not given any treatment.

Pre-treatment rectal faecal samples were collected from each animal and treated respectively according to group, while the animals from control group were identified and no treatment was given. After

10 to 14 days post-treatment, rectal faecal samples were collected again for faecal egg count (Anon., 1986). Faecal culture method was conducted to identify the species of nematode in each farm. The faecal culture method was done by pooling all the pre-treatment samples and culturing it for 5 to 7 days. Identification of the species of larvae using the identification keys by Manual of Veterinary Parasitological Laboratory Techniques (1971).

The data were analysed based on the on the calculations outlined by Coles *et al.* (1992). Resistance to a particular anthelmintic was considered to be present if the reduction in faecal egg count was less than 95% and the 95% lower confidence limit was less than 90%. If only one of the two criteria was met, the status was considered as suspected resistance. If the reduction in faecal egg count was more than 95% and the lower confidence limit was more than 90%, the status was considered as susceptible .

Standard questionnaires were used to interview the management of the farms on deworming history, farm management, feed and water source and mortality records.

## RESULTS

### Animals and Management

Information on the management of the animals in the farms are summarised in Table 1.

All the farms involved in this study are commercial farms breeding goats for meat purposes. The animals were crossbred Boer and Katjang, as the farmers kept various breeds together in their farm since they had started farming. The farms had a population of between 50 and 150 heads per farm. Two farms practiced semi intensive management and other 4 farms practiced free grazing management. The animals grazed around 4 to 5 hours per day and kept in raised floor and wooden sheds during night. Four of farms provided commercial concentrate feed supplement while in the shed. Animals in the shed also were given water ad libitum, salt or mineral blocks. Their mortality rate was between 10 to 40% a year due to the animals were free grazing and the farm management. Five farms (Farm A, C, D, E and F) were dewormed once or twice a year with ivermectin, and benzimidazole. Only one farm (Farm B) was never dewormed. However, the length of time these drugs have been used in the farm was unclear as no records were kept. According to the district veterinary department which institutes treatment, the common drugs administered to all small ruminant farms are from the Benzimidazoles group as it was economical.

### Anthelmintic resistance

Farm A, C and D shows severe resistance to all drug groups Benzimidazole (25%, -12%, 92%), Imidazothiazoles (33%, 93%, 97%), Salicylanilides (33%, -5%, -18%)

and Macrocytic Lactones (94%, -63%, 62%). Farm B and F shows resistance to 3 type of drugs Benzimidazole (-46%, 64%), Salicylanilides (54%,4%) and Macrocytic Lactones (-6%, 87% except Imidazothiazoles (97%, 96%). Farm E was resistant to one drug and susceptible to Imidazothiazoles, Macrocytic Lactones and Closantel (Table 2). The mean egg counts for each treatment group are given in Table 3.

Pre-treatment faecal culture from Farm A indicated that *Haemonchus contortus* is the predominant species (96%) followed by *Trichostrongylus* sp (3%) and *Cooperia* sp (1%). Post-treatment faecal cultures from Farm A showed resistance of *Haemonchus contortus*, sp., *Trichostrongylus* sp. and *Cooperia* sp. to Benzimidazole, Levamisole, Macrocytic Lactones (Ivermectin) and Closantel *Haemonchus contortus* is predominant species (50%) followed by *Oesophagostomum* sp. (38%) and *Trichostrongylus* sp. (12%) in Farm B. However, post treatment faecal cultures in Farm B showed resistance of *Haemonchus contortus*, *Oesophagostomum* sp., *Trichostrongylus* sp. and *Cooperia* sp. to Benzimidazole, Levamisole and Closantel. Macrocytic Lactones is the effective drug for Farm B.

Pre-treatment faecal culture from Farm C showed *Trichostrongylus* sp. is the predominant species (74%) followed by *Haemonchus contortus* (26%). Farm C showed that *Haemonchus contortus* and *Trichostrongylus* sp. is resistant to

all 4 drugs. The faecal cultures for Farm D indicated *Trichostrongylus* sp. (56%) was most common and followed by *Haemonchus contortus* (44%). For the post-treatment faecal cultures from Farm D, *Haemonchus contortus*, *Trichostrongylus* sp. were resistant to Benzimidazole, Imidazothiazoles, Salicylanilides and Macrocytic Lactones.

Pre-treatment faecal culture from Farm E indicated that *Trichostrongylus* sp is the predominant species (67%) followed by *Haemonchus contortus* (22%) and *Oesophagostomum* sp. (11%). Post-treatment faecal cultures from Farm E showed resistance of *Haemonchus contortus*, *Trichostrongylus* sp. and *Oesophagostomum* sp. to Benzimidazole whereas Closantel, Levamisole and Macrocytic Lactones is the effective drug for Farm E.

Pre-treatment faecal culture from Farm F showed *Haemonchus contortus* as the predominant species (48%) followed by *Trichostrongylus* sp. (28%) and *Oesophagostomum* sp. (24%). Post-treatment faecal cultures from Farm F showed resistance of *Haemonchus contortus*, sp., *Trichostrongylus* sp. and *Oesophagostomum* sp. to 3 drugs Benzimidazole, Macrocytic Lactones and Closantel. The effective drug for Farm F is Levamisole (Table 4).

## DISCUSSION

In Malaysia, anthelmintic resistance was suspected in the 1980's as small

**Table 1:** Summary of farm management in the 6 farms (Farms A-F) involved in this study

	Number of animals	Breed of animals	Management type	Feed	Grazing area	Mortality rate	Deworming frequency	Drug used
<b>Farm A</b>	60	Boer Mix	Semi-intensive	Grass when grazing and pellets while penned	Free roaming at oil palm plantation, always on the same area	33% a month	1/year	Ivermectin
<b>Farm B</b>	90	Boer Mix	Semi-intensive	Only grazing	Free roaming, always on the same area	13-40% a year	0	Animals were never dewormed
<b>Farm C</b>	140	Kaijang crosses	Free grazing	Grass when grazing, pellet	Free roaming under oil palm plantation	33% a month	2x/year	Benzimidazole
<b>Farm D</b>	55	Boer	Free grazing	Only grazing	Free roaming under oil palm plantation	23% a month	2x/year	Benzimidazole
<b>Farm E</b>	65	Boer Mix	Free grazing	Grass when grazing and pellets while penned	Free roaming under oil palm plantation	10% a month	2x/year	Benzimidazole
<b>Farm F</b>	100	Boer Mix	Free grazing	Grass when grazing and pellets while penned	Free roaming under oil palm plantation	10% a month	When Needed	Benzimidazole

**Table 2:** Anthelmintic resistance status on six smallholder farms (Farms A-F) .

	FECRT Percentage			
	Bz	Leva	ML	Clo
<b>Farm A</b>	25%	33%	94%	33%
<b>Farm B</b>	-46%	<b>97%</b>	-6%	54%
<b>Farm C</b>	-12%	93%	-63%	-5%
<b>Farm D</b>	92%	47%	62%	-18%
<b>Farm E</b>	95%	<b>99%</b>	<b>98%</b>	<b>96%</b>
<b>Farm F</b>	64%	<b>96%</b>	87%	4%

Note: FECR<50%-critical resistance; FECR 50% to 90%-severe resistance; FECR 91% to 95%- moderate resistance;>95% susceptible. Bold values indicate reduction greater than 95%. Bz = Benzimidazole, Leva = Levamisole, ML = Macrocylic Lactones (Ivermectin), Clo = Closantel (flukiver)

**Table 3:** Pre-treatment and post-treatment of mean faecal egg count (±SE) from treatment and control groups for each farm (Farms A-F)

		Mean Faecal egg count (epg)					
		Control	Bz	Leva	ML	Clo	
<b>Farm A</b>	Pre-treatment	625	2729	1775	2538	843	
	Post-treatment	1550	2267	43	1643	713	
<b>Farm B</b>	Pre-treatment	1010	937	1175	1362	3787	
	Post-treatment	463	350	312	28	1842	
<b>Farm C</b>	Pre-treatment	1010	890	333	1386	1970	
	Post-treatment	1070	1200	80	1743	1122	
<b>Farm D</b>	Pre-treatment	1200	1057	400	700	1271	
	Post-treatment	750	57	400	288	883	
<b>Farm E</b>	Pre-treatment	2617	800	600	1375	388	
	Post-treatment	117	125	29	43	100	
<b>Farm F</b>	Pre-treatment	2382	1929	1700	1738	2138	
	Post-treatment	1227	857	86	320	2288	

**Table 4:** Prevalence of third stage larvae (%), pre-treatment and post-treatment for each farm (Farms A-F ) from treatment and control groups.

			Control	Bz	Imid	ML	Clo
Farm A	Pre-treatment	<i>H. contortus</i> (96%), <i>Trichostrongylus</i> sp. (3%), <i>Cooperia</i> sp. (1%)					
	Post-treatment	<i>H. contortus</i>	84%	86%	97%	76%	86%
		<i>Trichostrongylus</i> sp.	11%	1%	3%	19%	1%
		<i>Cooperia</i> sp.	5%	13%	–	5%	13%
Farm B	Pre-treatment	<i>H. contortus</i> (50%), <i>Trichostrongylus</i> sp. (12%), <i>Oesophagostomum</i> sp. (38%)					
	Post-treatment	<i>H. contortus</i>	47%	50%	6%	84%	33%
		<i>Trichostrongylus</i> sp.	22%	49%	28%	16%	43%
		<i>Cooperia</i> sp.	2%	1%	1%	–	3%
		<i>Oesophagostomum</i> sp.	–	–	65%	–	21%
Farm C	Pre-treatment	<i>H. contortus</i> (26%), <i>Trichostrongylus</i> sp. (74%)					
	Post-treatment	<i>H. contortus</i>	28%	30%	20%	39%	17%
		<i>Trichostrongylus</i> sp.	72%	72%	80%	61%	83%
Farm D	Pre-treatment	<i>H. contortus</i> (44%), <i>Trichostrongylus</i> sp. (56%)					
	Post-treatment	<i>H. contortus</i>	44%	50%	0%	22%	28%
		<i>Trichostrongylus</i> sp.	56%	37%	100%	10%	72%
Farm E	Pre-treatment	<i>H. contortus</i> (22%), <i>Trichostrongylus</i> sp. (67%), <i>Oesophagostomum</i> sp. (11%)					
	Post-treatment	<i>H. contortus</i>	86%	18%	13%	1%	19%
		<i>Trichostrongylus</i> sp.	14%	3%	31%	2%	3%
		<i>Oesophagostomum</i> sp.	–	–	–	–	2%
Farm F	Pre-treatment	<i>H. contortus</i> (48%), <i>Trichostrongylus</i> sp. (28%), <i>Oesophagostomum</i> sp. (24%)					
	Post-treatment	<i>H. contortus</i>	31%	35%	20%	63%	25%
		<i>Trichostrongylus</i> sp.	16%	65%	76%	16%	25%
		<i>Oesophagostomum</i> sp.	–	–	4%	–	6%

Bz = Benzimidazole, Leva = Levamisole, Imid = Imidazothiazoles, Clo = Closantel

ruminant farmers usually complained of ineffective drugs and has been rapidly escalating. With an increasing trend to import sheep and goats from Australia, South Africa and South America where anthelmintic resistance is common, it is not surprising that the resistant worm populations may have been imported too. Repeated treatment of local animals with anthelmintics selects parasites that have innate or acquired resistance to the drugs. Ultimately treatment becomes ineffective because a large proportion of the parasites in the population are resistant. It is now common for sheep to harbour at least one parasite species that is resistant to the major drench groups. It is becoming disturbingly common for worms to be resistant to anthelmintics in all of the major drench groups, including some combinations of unrelated drugs. Drench resistance is common and often highly developed in goats. Only a limited range of anthelmintics are registered for use in goats. Due to their altered drug metabolism goats may require higher dose rates than recommended for sheep (Sani & Gray, 2004).

Traditionally 4 drug groups, namely benzimidazole (BZ), imidazothiazoles (Leva) (LEV), Macrocytic Lactones (IVM) were used control worms in small ruminants. However, with recent extensive spread of resistance in these chemical groups, other drenches including naphthalophos (NAP, Rametin TM), and its combinations with BZ and/or LEV, closantel (CLS), and other MLs such as

abamectin, and moxidectin should be considered (Sheep101.info. 2014). Although some may recommend an increased drug dose to combat anthelmintic resistance, in practice, only double dose LEV or double dose LEV plus single BZ have been shown to be effective where the single dose is not effective. However, it is emphasized that the use of double dose rate or combinations of anthelmintics are not general recommendations. They are used to obtain a prompt answer to possibly a serious and immediate problem on specific farms, and now are infrequently used (Vatta & Lindberg, 2006).

In Malaysia, farmer awareness on anthelmintic resistance is encouraged and various other means of worm control are advocated such as rotational grazing, use of herbs or cut and carry method of feeding instead of grazing permanent pastures under rubber or oil palm plantations. It is vital that farmers are aware of the dangers of importing new animals into their farm. With the diagnosis of anthelmintic resistance, farmers must practice some measure of biosafety and security to prevent resistant worm populations from establishing in their farm. Conducting this study will increase farmer awareness on the pitfalls of indiscriminate purchases of animals from unknown sources.

## REFERENCES

1. Anonymous, 1986 Manual of Veterinary Parasitological Laboratory Techniques. Reference Book 418. Third Edition . Ministry of Agriculture, Fisheries and Food. London. p160.



2. Chandrawathani P, Adnan M, Jamnah O (1998). The efficacy of moxidectin against trichostrongylid infection in sheep. *Journal of Bioscience* **9**: 112-113.
3. Chandrawathani P, Adnan M, Waller PJ (1999). Anthelmintic resistance in sheep and goat farms in Peninsular Malaysia. *Veterinary Parasitology* **82**: 305-310.
4. Chandrawathani P, Jamnah O, Chaeh TS, Adnan M, Rajamanickam C (1996). Evaluation of Closantel against gastrointestinal nematodes of sheep. *Journal of Bioscience* **7**: 83-86
5. Chandrawathani P, Premaalatha B, Nurulaini R, Erwanas AI, Zaini CM, *et al.* (2013). Severe anthelmintic resistance in two free grazing small holder goat farms in Malaysia. *J Veterinar Sci Technol* **4**: 137. doi:10.4172/2157-7579.1000137
6. Coles GC, Bauer C, Borgsteede FHM, Geerts S, Klei TR, *et al.* (1992). World Association for the Advancement of Veterinary Parasitology (WAAVP) methods for the detection of anthelmintic resistance in nematodes of veterinary importance. *Veterinary Parasitology* **44**: 35-44.
7. Dorny P, Claerebout E, Vercruysse J, Sani R, Jalila A (1994). Anthelmintic resistance in goats in peninsular Malaysia. *Vet Parasitol* **55**: 327-342.
8. Fatimah I, Ranjit KD, Jainudeen MR (1985). Causes of mortality of Dorset Horn Sheep in Malaysia. *Kajian Veterinar* **17**: 62-67.
9. Khadijah S, Rahman WA, Chandrawathani P, Waller PJ, Vasuge M, *et al.* (2006) Small Ruminants on Private Farms in Peninsular Malaysia: Nematode Resistance to Anthelmintic. *Journal Veterinary Malaysia* **18**: 29-32.
10. Khadijah S, Rahman WA, Chandrawathani P, Waller PJ, Vasuge M, *et al.* (2006) Nematode Anthelmintic Resistance in Government Small Ruminants Farms in Peninsular Malaysia. *Journal Veterinary Malaysia* **18**: 1-5
11. Rahman WA (1993). An assessment of thiabendazole resistant nematodes in some smallholder goat farms of Malaysia using the egg hatch assay method. *Veterinary Parasitology* **51**: 159-161.
12. Rahman WA (1994). Resistance to benzimidazole anthelmintic by *Haemonchus contortus* in goats in Peninsular Malaysia. *Veterinary Parasitology* **55**:155-157
13. Sani RA and Rajamanickam C (1990). Gastrointestinal parasitism in small ruminants in Malaysia. In: *ACIAR Proceedings* Np.74. Le Jambre and Knox (Ed.).Bogor, Indonesia. pp.98-100
14. Sani RA, Adnan M, Cheah TS and Chandrawathani P (2004). Worm control for small ruminants in Tropical Asia. *Australia Centre for International Agriculture Research Monograph* 113.
15. Sani RA and Gra GD (2004). Worm control in small ruminants in Southeast Asia. In: *Worm control for small ruminants in Tropical Asia*. ACIAR Monograph 113. pp. 3-21.
16. Sivaraj S, Dorny P, Vercruysse J and Pandey VS (1994). Multiple and multigeneric anthelmintic resistance on a sheep farm in Malaysia. *Veterinary Parasitology* **55**:159-165
17. Sheep101.info. 2014. Sheep 201: Internal parasite (worm) control. [online] Available at: <http://www.sheep101.info/201/parasite.htm>
18. Vatta AF and Lindberg ALE (2006). Managing anthelmintic resistance in small ruminant livestock of resource-poor farmers in South Africa. *Journal of the South African Veterinary Association*. (2006) **77**(1): 2-8 (En.). Onderstepoort Veterinary Institute, Private Bag X05, Onderstepoort, 0110 South Africa.